

# Journal of Drug Delivery and Biotherapeutics

Journal homepage: <u>https://sennosbiotech.com/JDDB/1</u>



# **Review Article**

# Liver X Receptor: A Potential Target for Hypertension

Dipti Patila:

<sup>a\*,</sup> Department of Pharmacology, SVKM's Dr. Bhanuben Nanavati College of Pharmacy, Vile Parle

## ARTICLEINFO

#### ABSTRACT

The Liver X receptor (LXR) emerges as a key player in the intricate relationship between hypertension and cardiovascular health. According to recent research, LXRs bind to a non-canonical response area in the renin promoter to regulate the transcription of the renin gene, which may have an impact on blood pressure regulation. LXR agonists have also demonstrated potential in lowering the rise in blood pressure brought on by Angiotensin II (Ang II), in part due to their suppression of AT1 and AT2 receptor gene expression. LXRs may potentially affect hypertension by influencing the amounts of TNF and NF-κB. Furthermore, nitrite levels are raised by LXR agonists in both normotensive and hypertensive states. These findings demonstrate how LXR can be targeted at different phases of the management of hypertension, which makes it a compelling therapeutic target for additional research in the area of cardiovascular health

Keywords: Hypertension; Liver X Receptor; Cardiovascular disease; Renin-angiotensin-aldosterone system

# **Corresponding Author:**

Dipti Patil

Department of Pharmacology, SVKM's Dr. Bhanuben Nanavati College of Pharmacy, Vile Parle

E-mail addresses:

Received date: 007-Mar-2024 Revised date: 18-Mar-2024, Accepted date: 18-Apr-2024

Crossref DOI: https://doi.org/10.61920/jddb.v1i01.26

© 2024 Sennos Biotech All rights reserved

#### 1. Introduction

Hypertension, defined as an increase in systolic, diastolic, or both blood pressures above normal ranges is widespread in both industrialized and developing countries, & its prevalence rises with age [1]. When the blood's long-term force against the artery walls is high enough to cause health issues like heart disease, it is referred to as hypertension or high blood pressure. The likelihood of developing additional health issues including heart disease, a heart attack, or a stroke increase with blood pressure [2].

The amount of blood your heart pumps, and the amount of resistance given by the arteries to the blood flow, influence your blood pressure. Your arteries become narrower as your heart pumps more blood pressure is measured in millimeters of mercury (mm Hg). Hypertension is the most common preventable risk factor for cardiovascular disease (CVD; including coronary heart disease, heart failure, stroke, myocardial infarction, atrial fibrillation, and peripheral artery disease), chronic kidney disease (CKD) is the greatest cause of death and disability worldwide, and it is linked to cognitive impairment [1] High blood pressure is still the greatest cause of mortality and disability [2] and it is expected to remain so in 2040 [3].

According to the 2003 Guidelines hypertensive blood pressure should be more than 140/90 mm Hg and according to 2017 Guidelines: hypertensive blood pressure should be more than 120/80mm Hg.

## **1.1 Types of Hypertensions**

#### Primary (Essential) Hypertension

In 90–95% of instances, primary hypertension is defined as high blood pressure brought on by a nonspecific lifestyle and hereditary factors. Risk factors include high body weight, smoking, drinking alcohol, and eating a diet high in sodium.

#### **Secondary Hypertension**

Secondary hypertension is considered as high BP caused because of an obvious cause like chronic renal disease, constriction of kidney arteries, a hormonal problem, or use of contraceptive pills. Secondary BP contributes to the remaining 5–10% of cases.

## 1.2 Pathophysiology of Hypertension

#### 1.2.1 BP regulation

This includes cardiac output, or the amount of blood the heart pumps out each minute, blood volume, and arterial tone, which is controlled by intravascular volume and neurohumoral systems. The sympathetic nervous system (SNS), the immunological system, natriuretic peptides, the endothelium, and the reninangiotensin-aldosterone system (RAAS) are all involved in regulating physiological blood pressure. The failure or disruption of one or more of these factors may lead to elevated mean blood pressure, fluctuating blood pressure, or both. These conditions may eventually cause organ damage (such as left ventricular hypertrophy), chronic kidney disease, and cardiovascular disease [4].

#### 1.2.2 Sodium Homeostasis

High blood Sodium content promotes water retention which in turn increases the Blood volume and blood pressure. Change in Systolic Blood pressure of a minimum of 10 mmHg under a few hours of ingestion of salt can give salt sensitivity. Individuals who are salt-sensitive have underlying endothelial dysfunction as a result of genetic or environmental factors. Because of their high salt load-induced overproduction of transforming growth factor (TGF) and low levels of accessible nitric oxide, these patients are more susceptible to oxidative stress and fibrosis. Even in those who are resistant to salt, a prolonged high-salt diet can cause endothelial dysfunction and alter gut flora, which can raise salt sensitivity and cause hypertension [5]. Consuming salt stimulates T helper cells, which in turn promotes autoimmunity [5].

## 1.2.3 Renin Angiotensin Aldosterone System

An important part of the pathogenesis of hypertension is the role of the RAAS in regulating blood pressure. In addition to playing a significant role in controlling blood pressure, it is in charge of Na+ retention, pressure natriuresis (which is defined as decreased Na+ reabsorption and increased Na+ excretion). salt sensitivity, vasoconstriction, endothelial dysfunction, and vascular damage [4]. It controls the kidney's pressure-volume equilibrium. Renin and its precursor pro-renin are produced and stored by the kidney's juxtaglomerular cells, which release them in response to external stimuli. Renin cleaves angiotensinogen to produce angiotensin I. The RAAS produces angiotensin II, which is what causes hypertension.

## **1.2.4 Natriuretic peptides**

ANP and BNP, the atrial and brain natriuretic peptides, regulate hypertension and salt sensitivity, respectively. Their significant natriuretic and vasodilator properties enable the maintenance of blood pressure and Na+ homeostasis during Na+ loading [6]. ANP and BNP are released in response to a Na+ load through atrial and ventricular stretch. This results in systemic vasodilation, a drop in plasma volume (from fluid transfers from the intravascular to the interstitial compartment), and a reduction in blood pressure [7]. Direct effects include decreased activity of the sodium-glucose co-transporter and Na+-K+-ATPase in the proximal tubule, as well as inhibition of the epithelial sodium channel in the distal nephron. Two indirect effects

are inhibition of aldosterone release and renin. A shortage of natriuretic peptides exacerbates hypertension. Corin is a serine protease that is mainly expressed in the heart and is responsible for converting the ANP and BNP precursors pro-ANP and pro-BNP to their active forms. Corin deficiency has been linked to volume overload, cardiac failure, and salt-sensitive hypertension [8]. The lack of natriuretic peptide is indeed linked to insulin resistance and type 2 diabetes. Obesity has been associated with natriuretic peptide shortage, which is thought to be caused by adipose tissue overexpression of the natriuretic peptide scavenger receptor NPR-C [9].

### 1.2.5 Sympathetic Nervous System

The SNS is more active in people with hypertension than in people who are not hypertensive. It is also more active in people who are obese, in men than in women, and in people who have advanced renal disease [10]. Identified indicators of sympathetic overactivity in normotensive people with a familial history of hypertension, such as elevated catecholamine spillover and sural nerve activity measured by microneurography [11]. Microneurography measurements of greater sympathetic activity are associated with a higher degree of hypertension [12]. Higher sympathetic activity is seen in patients with obesity-related hypertension, metabolic syndrome, and hypertension made worse by heart failure or renal disease [13].

## 1.2.6 Endothelial dysfunction

Vascular endothelial cells are important in cardiovascular regulation because they produce a variety of powerful local vasoactive agents, including the vasodilator chemical nitric oxide and the vasoconstrictor peptide endothelin. In humans, essential hypertension has been linked to endothelial dysfunction.

An appealing therapeutic strategy for trying to reduce some of the major consequences of hypertension is endothelial function modification. The generation of nitric oxide appears to be restored by clinically successful antihypertensive medication, although endothelium-dependent vascular relaxation and the vascular response to endothelial agonists do not appear to be restored. This suggests that once the hypertension process is begun, such endothelial damage is primary and irreversible.

## 2. Liver X Receptor

The liver X receptors (LXR) receptors belong to the nuclear receptor superfamily of ligand-activated transcription factors. Nuclear hormone receptors are a group of transcription regulators that play a role in a variety of physiological processes including cell proliferation, death, cancer, and angiogenesis. Because their natural ligands were unknown when they were discovered in the mid-1990s, they were dubbed "orphan receptors."[14][15][16]. Soon later, oxysterols and cholesterol metabolites were discovered to be endogenous LXR activators, and LXRs were "deorphanized."

Two isoforms of LXR, LXRbeta (NR1H2) and LXRalpha (NR1H3), combine to form heterodimers with retinoid X receptor (RXR). LXRs are intriguing therapeutic targets for the treatment of atherosclerotic and metabolic diseases since they have been shown to be critical regulators of inflammation, lipid and glucose metabolism, and cholesterol homeostasis [17].

LXRs are connected to LXR response elements (LXREs) in target gene regulatory regions and form obligatory heterodimers with the retinoid X receptor (RXR) in the nucleus. Though it is expressed in every part of the body, metabolically active systems such the liver, adipose tissue, and macrophages, as well as the heart, skeletal muscle, kidney, and lung, are where LXR is most commonly found [18].

When LXR target genes are activated, LXR binds to the LXR response element in the promoter region to regulate the expression of those genes. These genes are involved in the metabolism of glucose, lipids, fatty acids, and cholesterol. These consist of fatty acid synthase (FAS), lipoprotein lipase, sterol regulatory element-binding protein-1c (SREBP-1c), apolipoprotein E (ApoE), cholesteryl ester transfer protein, ATP-binding cassette (ABC) transporter isoforms A1 (ABCA1) and G1 (ABCG1), and glucose.

LXR is an oxysterol sensor with LXR $\alpha$  which contains 447 amino acids and LXR $\beta$  which contains 461 [19] amino acids in its structure. The four functional domains that make up the LXR molecules are: the hydrophobic ligand-binding domain (LBD), which is necessary for ligand binding and receptor dimerization; the amino-terminal ligandindependent activation function domain (AF-1), which can stimulate transcription in the absence of a ligand; and the DNA-binding domain (DBD) with two zinc fingers.

LXR $\alpha$  and LXR $\beta$  have 75.6 percent and 74% sequence identity in their DBD and LBD regions, respectively [20]. The LBD of LXR has a three-layered -helical sandwich shape with ten  $\alpha$ -helices, which is similar to all known nuclear receptors [21]. LXR/RXR is a "permissive heterodimer" that can be activated by either an LXR agonist or a particular RXR ligand called 9-cis retinoic acid. The LXR/RXR complex binds specifically to an LXR responsive element (LXRE), which is made up of two direct repeats of hexameric nucleotides, AGGTCA, separated by four or one nucleotide(s) in

the promoter region of target genes (DR4 or DR1) [22].

A large number of LXR target genes have been found, including fatty acid synthase (FAS), ATP binding cassette (ABC), transporter isoforms A1, G1, G5, and G8, apolipoprotein E (ApoE), cholesteryl ester transfer protein (CETP), cytochrome P450 isoform 7A1 (CYP7A1)cholesterol 7a-hydroxylase, and carbohydrate regulatory element binding protein (ChREBP) [23]. LXRs are connected to LXR response elements (LXREs) in target gene regulatory regions and form obligatory heterodimers with the retinoid X receptor (RXR) in the nucleus. The LXR/RXR complex undergoes a conformational shift upon binding with either natural or synthetic ligands, such as T0901317 (T09) and GW3965, which subsequently activates target gene transcription.

## 2.1 Function

There are different functions of LXR in the body such as cholesterol metabolism regulation, boosting insulin secretion, and enhancing insulin sensitivity. Also, LXRs are anti-inflammatory and antiautoimmune receptors. **LXRs** are antiatherosclerotic [24] also cause renin secretion regulation [25], and inhibit the formation of amyloid plaque in the central nervous system [26], LXRs also have Gonadal function and steroidogenesis control in the gonads and adrenals [27]. Changes in endogenous LXR activity are seen in a variety of clinical situations, including atherosclerosis, cancer, neurological illnesses such as multiple sclerosis, Alzheimer's disease, Parkinson's disease, arthritis, and skin diseases.

# 2.2 Liver X receptor Agonists

## 2.2.1 Endogenous agonist

Oxysterol has been discovered to be an endogenous ligand for LXRs in mammals, with two separate

types of ligands that activate LXRs [28][29]. 20(S)-, 22(R)-, 24(S)-, 25-, and 27-hydroxy cholesterol, as well as 24(S), 25- epoxy cholesterols, are oxidized derivatives of oxysterol. The intermediates of cholesterol production are the second class of LXR-activating oxysterols. The oxysterol 24(S), 25- epoxycholesterol is formed in a shunt of the mevalonate pathway.

24(S)-hydroxycholesterol is a kind of 24(S)hydroxycholesterol (cerebrosterol) is abundant in the brain. 24-hydroxylase is responsible for its production. It has been shown to be a good activator of the LXR-regulated gene ABCA1 [30]. The P450 enzyme sterol 27-hydroxylase produces 27hydroxycholesterol 27from cholesterol. hydroxylase then oxidises it to aldehyde and carboxylic acid (cholestenoic acid). The ligands for LXRs are 27-hydroxycholesterol and cholestenoic acid [31][32]. 25-hydroxycholesterol is a strong regulator of LXR-mediated pathways, is synthesised by 25-hydroxylase. It also has an effect on the expression of LXR-dependent genes such as LPL, ABCG5, and ABCG8 [32]. 24(S) 25epoxycholesterol produced in the mevalonate pathway's shunt. The expression of LXR target genes ABCA1 is reduced when 24(S),25epoxycholesterol is removed [33]. Desmosterol and zymosterol are two more intermediates that activate the LXR [34].

# 2.2.2 Natural agonists

Fucosterol, which is abundant in marine algae, exhibits hypocholesterolemic properties and raises plasma HDL activity. In HEK–293 cells, fucosterol greatly increased the transactivation of both LXR $\alpha$ and LXR $\beta$  [35]. Podocarpic acid is the LXR agonist produced from plant resins that is non-steroidal [36]. Experiments using LXR scintillation proximal binding demonstrated that compounds of podocarpic acid, including its dimer anhydride and imides, bind to both LXRα and LXRβ. Cell-based transactivation experiments on HEK-293 cells showed that the anhydride dimer had an EC50 value of 1 nM against both receptors, with peak induction for the  $\beta$  and  $\beta$ LXR receptors being 50- and 8-fold, respectively. In hamsters, the more stable and powerful imide raises cholesterol 28% total plasma by while simultaneously raising HDL cholesterol by 22% and lowering LDL cholesterol by 11%. Similar findings were shown in mice with HDL cholesterol levels that were boosted by 19% [37]. Cyanidin, a flavonoid present in a variety of fruits and vegetables has been shown to affect cellular lipid metabolism. LXRa transactivation was increased by 32% (at 50 M), 59 percent (at 100 M), and 33% (at 100 M) in response to cyanidin. With an EC50 of 3.48 M of LXRa and 125.2 M of LXRB, cyanidin activates the LXR. LXR-sensitive genes such as ABCA1, SREBP-1c, and ABCG5 were likewise activated by cyanidin. It also lowered cellular TG contents [38].

Cineole, a small fragrance component found in teas and plants, has been found to promote LXR transactivation. Cineole treatment of CHO-K1 cells increased LXR transactivation by more than 75% and LXR transactivation by more than 100%. Cineole reduced cellular cholesterol levels in RAW 264.7 macrophages. The mRNA expression of the LXR-responsive genes was similarly dramatically enhanced by cineole. Surprisingly, LXR-sensitive SREBP-1c, and SCD-1 were genes FAS, significantly downregulated in hepatocytes activated by cineole. Cineole acts as a partial agonist, activating LXRs selectively without promoting hepatic lipogenesis [39]

# 2.2.3 Synthetic Agonist

The synthetic ligands T0314407 and T0901317 both increased LXR transcriptional activity. T0901317 as a nonsteroidal LXRα ligand with exceptional potency and selectivity. While T0314407 and T0901317 bind to LXR $\alpha$  with EC50 values of 100 and 20 nM, respectively, they were substantially more powerful and had equivalent efficacy to 24,25-EC.

In a cell-free ligand-sensing test, GW3965 is a steroid receptor coactivator-1 to human LXRalpha with an EC(50) of 125 nM. In cell-based reporter gene assays, it profiles as a complete agonist on hLXRalpha and hLXRbeta with EC(50) values of 190 and 30 nM, respectively. increased plasma HDL cholesterol concentrations by 30% [40][41] and enhanced the expression of the reverse cholesterol transporter ABCA1 in the small intestine and peripheral macrophages following oral treatment of 10 mg/kg to C57BL/6 mice.

LXR-623, a Synthetic ligand for LXRs that has shown potential in atherosclerosis animal models. LXR activation increased ABCA1 and ABCG1 expression in a dose-dependent manner. A population pharmacokinetic-pharmacodynamic study was used to further evaluate the influence of LXR-623 concentration on ABCA1 and ABCG1 expression, producing EC50 estimations of 526 ng/mL and 729 ng/mL, respectively [42][43].

AZ876 which is one of the novel high-affinity LXR agonists. LXR could thus be a potential target for antihypertrophic and antifibrotic treatments in the prevention of heart failure. The hypertrophic and profibrotic transforming growth factor  $\beta$  (TGF $\beta$ )-Smad2/3 signalling was further reduced by AZ876, which also repressed the up-regulation of genes linked to fibrosis and hypertrophy.AZ876 decreased insmooth muscle actin, inhibition of TGF and angiotensin II-induced fibroblast collagen synthesis, and modulation of the Myo fibroblastic marker. After receiving AZ876 therapy, liver weight and plasma triglycerides were unchanged. Comparing AZ876 to GW3965 (HY-10627), it is 25-fold and 2.5-fold more effective on human hLXR $\alpha$  and hLXR $\beta$ , respectively [44][45].

The partial LXR agonist BMS-779788 has IC50 values of 14 nM for LXR<sup>β</sup> and 68 nM for LXR<sup>α</sup>, respectively. The selective partial agonist LXR was found in human whole blood. When it comes to stimulating the ATP-binding transporters ABCA1 and ABCG1, BMS-779788 is quite effective [46]. BMS-779788 has an EC50=610 nM, which is comparable to its in vitro blood gene induction potency, when it comes to inducing LXR target genes in blood in vivo. BMS-779788 is 29- and 12fold less efficacious than the full agonist T0901317 in raising plasma triglycerides and LDL cholesterol, respectively. Similar outcomes are shown for plasma cholesteryl ester transfer protein and apolipoprotein B. BMS-779788 induces ABCA1 peripherally in mice, with no significant increase in plasma or hepatic triglycerides, indicating a better profile than a complete pan-agonist [47].

BMS-852927, LXRβ-selective agonist with 20% LXRa and 88% LXRB activity compared to a full pan agonist in transactivation assays. BMS-852927 has a binding affinity to both LXR $\alpha$  and LXR $\beta$  in a similar way. In cynomolgus monkeys and mice, XL041 (BMS-852927) has a very good efficacy profile. Pre-treatment of C57BL/6J mice with XL041 for 7 days causes a significant, dosedependent increase of cholesterol efflux in this system, with a maximum of 70% above vehicle in the initial efflux rate in the [48] 3 mg/kg/day dosing group. LDLR knockout (KO) mice produced similar results. A 12-week research on LDLR KO mice found that XL041 inhibited the progression of atherosclerosis. The dose response for inhibition of atherosclerosis is (0.1-3 mg/kg/day) [49].

## 2.3 Role of LXR in Hypertension

Hypertension damages the heart muscle by increasing hemodynamic afterload, which starts the growth of pathological hypertrophy. It also increases the risk of atherosclerosis and damages the arteries. An essential hormonal signalling mechanism in the control of blood pressure, fluid equilibrium, and systemic vascular resistance is the RAAS. Blood pressure modulation has been linked to LXRmediated RAAS regulation. LXR was originally discovered to be a transcription regulator of renin [50][51]. LXR signalling and the RAAS are connected, as evidenced by the fact that acute LXR agonist treatment raised renin mRNA levels in vivo and LXR-null animals were unable to upregulate renin in response to -adrenergic stress [50]. The LXR agonist T09 was found to reduce blood pressure increase in mice with chronic pressurevolume overload, but this effect was lost in mice lacking LXR cardiac LXR overexpression increases natriuretic peptide expression. LXR modulation of natriuretic peptides might therefore be an indirect technique for RAAS suppression; hence, LXRs limit RAAS activation and could be a viable target for lowering the heart's hemodynamic stress [52].

## 2.4 LXR effect on hypertension

## 2.4.1 AT1R expression

Angiotensin II (Ang II) is mediated by Ang II receptors, of which there are now two isoforms known to exist: type 1 receptor (AT1R) and type 2 receptor. Most of Ang II's typical effects, such as cell proliferation and vasoconstriction, are mediated through AT1R. The promotion of atherogenesis by Ang II and the elevation of atherogenesis by an AT1R are well established [53]. AT1R expression was likewise decreased by 22-(R)hydroxycholesterol, a well-known LXR ligand. T0901317 needed de novo protein synthesis to downregulate AT1R. LXR Agonist Reduced AT1R mRNA and Protein Expression At 6 hours after

incubation with T0901317, AT1R mRNA was reduced in a dose-dependent manner. T0901317 decreased AT1R protein levels, with a peak at 6 hours of incubation.

T0901317-Induced Downregulation of AT1R Expression Requires De Novo Protein Synthesis. T0901317-induced downregulation of AT1R expression was tested using CHX (10 g/mL, 1 hour), a protein synthesis inhibitor, to see if it was dependent on de novo protein synthesis. The expression of AT1R mRNA was unaffected by CHX T0901317induced AT1R mRNA alone. downregulation was, however, blocked by CHX. De novo protein synthesis is required for AT1R mRNA downregulation by LXR. p16 suppressed Sp1mediated gene transcription by suppressing cyclin A expression and phosphorylating Sp1 at the serine position. T0901317 (10 mol/L) enhanced p16 expression and decreased Sp1 phosphorylation at the serine residue, indicating that it boosted p16 expression and lowered Sp1 phosphorylation.

AT1R Downregulation Caused by T0901317 Reduced Angiotensin II Cellular Response Through AT1R, Ang II causes phosphorylation of ERK in VSMCs. When AT1R expression is maximally inhibited, ERK phosphorylation is dramatically reduced after 6 to 12 hours of preincubation with T0901317. However, after incubation with T0901317 for 6 to 12 hours, phorbol ester (100 nM) boosted ERK phosphorylation, indicating that T0901317 may not influence the ERK activation pathway (data not shown). As a result, AT1R downregulation resulted in a reduction in the cellular response to Ang II.

LXR-deficient (LXR-/) and wild-type (WT) C57Bl/6J mice were given T0901317 (T09, a selective synthetic LXR agonist) along with isoproterenol, a RAAS inducer, and then either ISO, T09, or both for seven days. Low-dose ISO treatment in WT mice increased renal renin mRNA, renin protein, and ACE protein; however, this increase was uncorrelated with elevated blood pressure. WT mice treated with both ISO and T09 exhibited decreased renal renin, ACE, and AT1R mRNA expression in comparison to mice treated with ISO alone. In addition, WT mice treated with ISO plus T09 showed lower cardiac ACE mRNA expression in their hearts than did those treated with ISO alone. Animals lacking LXR largely lacked transcriptional changes for renin, ACE, and AT1R.

A synthetic liver X receptor (LXR) agonist called GW3965 in Sprague-Dawley rats reduces angiotensin II-mediated pressor responses [54]. Synthetic LXR agonist GW3965 decreased vascular Ang II receptor gene expression and Ang IImediated blood pressure increases. These novel results identify LXRs as key players in blood pressure regulation, RAAS, and lipid metabolism.

## 2.4.2 Renin modulation

Both LXRs controlled renin transcription in vitro by binding to a noncanonical responsive region in the renin promoter. While LXR was a cAMP-activated factor, cAMP had the opposite effect on LXR. In vivo, LXRs colocalized with the renin promoter in juxtaglomerular cells, where LXR was especially concentrated. Renin is an aspartyl protease that catalyzes the first and rate-limiting stage in the renin-angiotensin cascade, the cleavage of angiotensinogen to angiotensin I. Specialized juxtaglomerular (JG) cells strategically placed in the afferent arterioles of kidney glomeruli express and release circulating renin. Renin regulates the rate at which angiotensin is produced, which is important for salt-volume balance and blood pressure management.

LXRs bind to the renin promoter and hence control the expression of the renin gene. The intracellular cAMP increase was followed by overexpression of renin mRNA in As4.1/LXR cells 6 hours after stimulation. Overexpression of LXR, on the other hand, was linked to greater basal levels of renin mRNA, whereas cAMP treatment was linked to a decrease in renin mRNA levels [51].

In JG cells in vivo, LXRs are expressed with a particular enrichment in LXR. LXR was expressed at equal quantities in both sorted and unsorted cells, whereas JG cells showed a substantial enrichment in LXR. As a result, independent methods revealed that both LXRs are expressed in JG cells' kidneys, with a particular enrichment in LXR.

## 2.4.3 Vasoreactivity

Reduced Ang II receptor gene expression in response to LXR medication, possibly explaining the reduced Ang II-induced vasopressor response. Rats given GW3965 are anticipated to exhibit reduced vasoreactivity in response to Ang II infusions due to AT1 receptor downregulation. It's important to note that Ang II-mediated blood pressure was shown to be lower at the 6–8-hour time intervals, which were characterized by a tendency towards decreased Ang II receptor gene expression [51]

GW3965 therapy over the previous week resulted in a significant reduction in the hypertensive rats' systolic blood pressure (p<0.05). In normotensive mice, GW3965 therapy raised plasma nitrite levels (p<0.05)

## 2.4.4 Nitrite levels

The impact of GW3965 on the levels of plasma nitrite. The plasma nitrite levels in the hypertensive group were constant. Nitrite levels were elevated by GW3965 treatment in rats with normal and hypertension. It was discovered that the LXR agonist improved TNF-impaired NO production in endothelial cells and reversed the TNF-induced downregulation of eNOS expression [55]. Plasma NO levels were raised in rats with normotension by GW3965 treatment. The beneficial effect on vessels may be attributed in part to the LXR agonist GW3965's effects on NO levels.

## 2.4.5 NF-κB and TNF-α

It has been demonstrated that the mesenteric artery of hypertensive rats expresses more NF-kB protein. TNF is a cytokine that promotes inflammation and is regulated by transcription It has been observed that hypertensive rats exhibit increased levels of NF- $\kappa$ B TNF- and NF- $\kappa$ B protein expression in the aorta [55]. GW3965 treatment brought the increased TNF-TNF-expression level in hypertensive rats down to the control level. The administration of GW3965 has an impact on TNF expression and NF- $\kappa B$  levels. In hypertensive rats, the expressions of TNF- (A, C) and NF-κB (A, B) were markedly elevated, and GW3965 treatment greatly reduced them. NF-kB expression in the aorta was increased in hypertensive rats (p<0.05). In hypertensive individuals, GW3965 treatment decreased NF-KB protein expression (p<0.05).

# 5. Conclusion

Hypertension is the leading cause of death around the world. The Liver X receptor is the key receptor that can be a relationship between hypertension and the LXR receptor, as we saw in the report. The discovery that LXRs regulate renin gene transcription by binding to a noncanonical response region in the renin promoter has led to speculation that LXRs may play a role in facilitating this crosstalk. Blood pressure rises caused by Ang II were decreased by using an LXR agonist. The downregulation of vascular AT1 and AT2 receptor gene expression was linked to decreased vascular responsiveness to Ang II treatment. AT1 receptor stimulation is principally responsible for changes in vascular function associated with RAAS activation. NF- $\kappa$ B expression increases in hypertensive rats, so LXR agonist acts on NF- $\kappa$ B and also TNF and decreases its level in hypertensive rats. Also, the LXR agonists increase nitrite levels in normotensive and also in hypertensive rats T0901317, a synthetic LXR agonist, inhibited AT1R mRNA and protein expression. So, targeting the liver X receptor at various stages can prove to be very useful with respect to hypertension. Thus, making a Liver X receptor a potential target for hypertension.

### Acknowledgment

I would like to thank the Department of Pharmacology, SVKM'S Dr. Bhanuben Nanavati College of Pharmacy, Vile Parle for the guidance and support.

### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Authorship contribution statement

**Dipti Patil**: Conceptualization, Writing – original draft.

#### Abbreviations

CVD: Cardiovascular diseases

- CKD: Chronic obstructive disease
- RAAS: Renin-angiotensin-aldosterone system
- SNS: Sympathetic nervous system
- TGF: Transforming growth factor
- ANP: Atrial natriuretic peptide
- BNP: brain natriuretic peptide
- NPR-C: natriuretic peptide scavenger receptor
- APA: Aminopeptidase
- ACE2: Angiotensin-converting enzyme 2
- NP: Natriuretic peptide
- CNP: C-type natriuretic peptide
- pGC: particulate guanylyl cyclase
- PK-G: protein kinase G
- MAPK: mitogen-activated protein kinase

PDE: phosphodiesterase DOCA : Deoxycorticosterone acetate LXR : Liver X receptor ABC: ATP-binding cassette FAS: Fatty acid synthase ApoE: Apolipoprotein E SREBP-1c: sterol regulatory element-binding protein-1c DBD: DNA-binding domain LBD: ligand-binding domain ISO: Isoproterenol

# REFERENCES

- [1] Oparil S, Acelajado MC, Bakris GL, Berlowitz DR, Cífková R, Dominiczak AF, et al. Hypertension. Nat Rev Dis Prim 2018 Jun 7;4(1):18014. Available from: http://www.nature.com/articles/nrdp201814
- [2] Forouzanfar MH, Afshin A, Alexander LT, Anderson HR, Bhutta ZA, Biryukov S, et al. Global, regional, and national comparative risk assessment of 79 behavioural, environmental and occupational, and metabolic risks or clusters of risks, 1990– 2015: a systematic analysis for the Global Burden of Disease Study 2015. Lancet . 2016 Oct;388(10053):1659–724. Available from:

https://linkinghub.elsevier.com/retrieve/pii/ S0140673616316798

[3] Foreman KJ, Marquez N, Dolgert A, Fukutaki K, Fullman N, McGaughey M, et al. Forecasting life expectancy, years of life lost, and all-cause and cause-specific mortality for 250 causes of death: reference and alternative scenarios for 2016–40 for 195 countries and territories. Lancet . 2018 Nov;392(10159):2052–90. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0140673618316945

- [4] Hall ME, Hall JE. Pathogenesis of Hypertension. In: Hypertension: A Companion to Braunwald's Heart Disease.
   Elsevier; 2018. p. 33–51. Available from: https://linkinghub.elsevier.com/retrieve/pii/ B9780323429733000056
- [5] Wilck N, Matus MG, Kearney SM, Olesen SW, Forslund K, Bartolomaeus H, et al. Salt-responsive gut commensal modulates TH17 axis and disease. Nature . 2017 Nov 15;551(7682):585–9. Available from: http://www.nature.com/articles/nature2462 8
- [6] Kerkelä R, Ulvila J, Magga J. Natriuretic Peptides in the Regulation of Cardiovascular Physiology and Metabolic Events. J Am Heart Assoc . 2015 Oct 27;4(10). Available from: https://www.ahajournals.org/doi/10.1161/J AHA.115.002423
- [7] Curry F-RE. Atrial natriuretic peptide: an essential physiological regulator of transvascular fluid, protein transport, and plasma volume. J Clin Invest . 2005 Jun 1;115(6):1458–61. Available from: http://www.jci.org/cgi/doi/10.1172/JCI254 17
- [8] Armaly Z, Assady S, Abassi Z. Corin. Curr Opin Nephrol Hypertens . 2013 Nov;22(6):713–22. Available from: https://journals.lww.com/00041552-201311000-00019
- Schlueter N, de Sterke A, Willmes DM,
   Spranger J, Jordan J, Birkenfeld AL.
   Metabolic actions of natriuretic peptides and therapeutic potential in the metabolic syndrome. Pharmacol Ther . 2014
   Oct;144(1):12–27. Available from:

https://linkinghub.elsevier.com/retrieve/pii/ S0163725814000953

- [10] Augustyniak RA, Picken MM, Leonard D, Zhou XJ, Zhang W, Victor RG. Sympathetic nerves and the progression of chronic kidney disease during 5/6 nephrectomy: Studies in sympathectomized rats. Clin Exp Pharmacol Physiol . 2010 Jan;37(1):12–8. Available from: https://onlinelibrary.wiley.com/doi/10.1111 /j.1440-1681.2009.05253.x
- [11] Grassi G, Mark A, Esler M. The Sympathetic Nervous System Alterations in Human Hypertension. Circ Res . 2015 Mar 13;116(6):976–90. Available from: https://www.ahajournals.org/doi/10.1161/C IRCRESAHA.116.303604
- [12] Grassi G, Cattaneo BM, Seravalle G, Lanfranchi A, Mancia G. Baroreflex Control of Sympathetic Nerve Activity in Essential and Secondary Hypertension. Hypertension . 1998 Jan;31(1):68–72. Available from: https://www.ahajournals.org/doi/10.1161/0 1.HYP.31.1.68
- Smith P. Relationship between central sympathetic activity and stages of human hypertension. Am J Hypertens . 2004 Mar;17(3):217–22. Available from: https://academic.oup.com/ajh/article-lookup/doi/10.1016/j.amjhyper.2003.10.01 0
- [14] Song C, Kokontis JM, Hiipakka RA, Liao S.
  Ubiquitous receptor: a receptor that modulates gene activation by retinoic acid and thyroid hormone receptors. Proc Natl Acad Sci . 1994 Nov 8;91(23):10809–13.
  Available from:

https://pnas.org/doi/full/10.1073/pnas.91.23 .10809

- [15] Apfel R, Benbrook D, Lernhardt E, Ortiz MA, Salbert G, Pfahl M. A novel orphan receptor specific for a subset of thyroid hormone-responsive elements and its interaction with the retinoid/thyroid hormone receptor subfamily. Mol Cell Biol . 1994 Oct;14(10):7025–35. Available from: http://mcb.asm.org/lookup/doi/10.1128/MC B.14.10.7025
- [16] Willy PJ, Umesono K, Ong ES, Evans RM, Heyman RA, Mangelsdorf DJ. LXR, a nuclear receptor that defines a distinct retinoid response pathway. Genes Dev . 1995 May 1;9(9):1033–45. Available from: http://genesdev.cshlp.org/lookup/doi/10.11 01/gad.9.9.1033
- [17] Tontonoz P, Mangelsdorf DJ. Liver X Receptor Signaling Pathways in Cardiovascular Disease. Mol Endocrinol .
  2003 Jun 1;17(6):985–93. Available from: https://academic.oup.com/mend/article/17/ 6/985/2747382
- Peet DJ, Janowski BA, Mangelsdorf DJ.
   The LXRs: a new class of oxysterol receptors. Curr Opin Genet Dev . 1998 Oct;8(5):571–5. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0959437X98800130
- [19] Shinar DM, Endo N, Rutledge SJ, Vogel R, Rodan GA, Schmidt A. NER, a new member of the gene family encoding the human steroid hormone nuclear receptor. Gene . 1994 Sep;147(2):273–6. Available from: https://linkinghub.elsevier.com/retrieve/pii/ 0378111994900809

- Tice CM, Noto PB, Fan KY, Zhuang L, Lala DS, Singh SB. The Medicinal Chemistry of Liver X Receptor (LXR) Modulators. J Med Chem . 2014 Sep 11;57(17):7182–205. Available from: https://pubs.acs.org/doi/10.1021/jm500442 z
- [21] Svensson S. Crystal structure of the heterodimeric complex of LXR and RXR ligand-binding domains in a fully agonistic conformation. EMBO J . 2003 Sep 15;22(18):4625–33. Available from: http://emboj.embopress.org/cgi/doi/10.1093 /emboj/cdg456
- [22] Hu X, Li S, Wu J, Xia C, Lala DS. Liver X Receptors Interact with Corepressors to Regulate Gene Expression. Mol Endocrinol
  . 2003 Jun 1;17(6):1019–26. Available from: https://academic.oup.com/mend/article/17/ 6/1019/2747398
- [23] Edwards PA, Kennedy MA, Mak PA. LXRs; Vascul Pharmacol . 2002 Apr;38(4):249–56. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S1537189102001751
- [24] Lee SD, Tontonoz P. Liver X receptors at the intersection of lipid metabolism and atherogenesis. Atherosclerosis . 2015 Sep;242(1):29–36. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0021915015300010
- [25] Pan L, Gross KW. Transcriptional Regulation of Renin. Hypertension . 2005 Jan;45(1):3–8. Available from: https://www.ahajournals.org/doi/10.1161/0 1.HYP.0000149717.55920.45

- [26] Sandoval-Hernández AG, Buitrago L, Moreno H, Cardona-Gómez GP, Arboleda G. Role of Liver X Receptor in AD Pathophysiology. Planel E, editor. PLoS One . 2015 Dec 31;10(12):e0145467. Available from: https://dx.plos.org/10.1371/journal.pone.01 45467
- [27] Cummins CL. Liver X receptors regulate adrenal cholesterol balance. J Clin Invest .
   2006 Jul 3;116(7):1902–12. Available from: http://www.jci.org/cgi/doi/10.1172/JCI284 00
- [28] Janowski BA, Willy PJ, Devi TR, Falck JR, Mangelsdorf DJ. An oxysterol signalling pathway mediated by the nuclear receptor LXRα. Nature . 1996 Oct;383(6602):728– 31. Available from: http://www.nature.com/articles/383728a0
- [29] Lehmann JM, Kliewer SA, Moore LB, Smith-Oliver TA, Oliver BB, Su J-L, et al. Activation of the Nuclear Receptor LXR by Oxysterols Defines a New Hormone Response Pathway. J Biol Chem . 1997 Feb;272(6):3137–40. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0021925819783550
- [30] BJÖRKHEM I. Crossing the barrier: oxysterols as cholesterol transporters and metabolic modulators in the brain. J Intern Med . 2006 Dec;260(6):493–508. Available from: https://onlinelibrary.wiley.com/doi/10.1111 /j.1365-2796.2006.01725.x
- [31]PikulevaIA.CHOLESTEROL-METABOLIZINGCYTOCHROMESP450.DrugMetabDispos.2006Apr;34(4):513–20.Availablefrom:

http://dmd.aspetjournals.org/lookup/doi/10. 1124/dmd.105.008789

- [32] Song C, Liao S. Cholestenoic Acid Is a Naturally Occurring Ligand for Liver X Receptor α\*\*This work was supported by NIH grants. Endocrinology . 2000 Nov 1;141(11):4180–4. Available from: https://academic.oup.com/endo/article/141/ 11/4180/2987854
- [33] Chen W, Chen G, Head DL, Mangelsdorf DJ, Russell DW. Enzymatic Reduction of Oxysterols Impairs LXR Signaling in Cultured Cells and the Livers of Mice. Cell Metab . 2007 Jan;5(1):73–9. Available from: https://linkinghub.elsevier.com/retrieve/pii/

https://linkinghub.elsevier.com/retrieve/pii/ S1550413106003950

 [34] Yang C, McDonald JG, Patel A, Zhang Y, Umetani M, Xu F, et al. Sterol Intermediates from Cholesterol Biosynthetic Pathway as Liver X Receptor Ligands. J Biol Chem .
 2006 Sep;281(38):27816–26. Available from:

> https://linkinghub.elsevier.com/retrieve/pii/ S0021925819580668

[35] Hoang M-H, Jia Y, Jun H, Lee JH, Lee BY, Lee S-J. Fucosterol Is a Selective Liver X Receptor Modulator That Regulates the Expression of Key Genes in Cholesterol Homeostasis in Macrophages, Hepatocytes, and Intestinal Cells. J Agric Food Chem . 2012 Nov 21;60(46):11567–75. Available from:

https://pubs.acs.org/doi/10.1021/jf3019084

[36] Sherwood IR, Short WF. 192. Podocarpic acid. Part I. J Chem Soc . 1938;1006.Available from: http://xlink.rsc.org/?DOI=jr9380001006

- [37] Singh SB, Ondeyka JG, Liu W, Chen S, Chen TS, Li X, et al. Discovery and development of dimeric podocarpic acid leads as potent agonists of liver X receptor with HDL cholesterol raising activity in mice and hamsters. Bioorg Med Chem Lett . 2005 Jun;15(11):2824–8. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0960894X05004221
- [38] Jia Y, Hoang MH, Jun H-J, Lee JH, Lee S-J. Cyanidin, a natural flavonoid, is an agonistic ligand for liver X receptor alpha and beta and reduces cellular lipid accumulation in macrophages and hepatocytes. Bioorg Med Chem Lett . 2013 Jul;23(14):4185–90. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0960894X13006252
- [39] Jun H, Hoang M-H, Yeo S-K, Jia Y, Lee S-J. Induction of ABCA1 and ABCG1 expression by the liver X receptor modulator cineole in macrophages. Bioorg Med Chem Lett . 2013 Jan;23(2):579–83. Available from:

https://linkinghub.elsevier.com/retrieve/pii/ S0960894X12014552

- [40] Guo D, Reinitz F, Youssef M, Hong C, Nathanson D, Akhavan D, et al. An LXR Agonist Promotes Glioblastoma Cell Death through Inhibition of an EGFR/AKT/SREBP-1/LDLR-Dependent Pathway. Cancer Discov 2011 Oct;1(5):442–56. Available from: http://cancerdiscovery.aacrjournals.org/loo kup/doi/10.1158/2159-8290.CD-11-0102
- [41] Spyridon M, Moraes LA, Jones CI, Sage T, Sasikumar P, Bucci G, et al. LXR as a novel antithrombotic target. Blood . 2011 May

26;117(21):5751–61. Available from: https://ashpublications.org/blood/article/11 7/21/5751/21424/LXR-as-a-novelantithrombotic-target

- [42] Villa GR, Hulce JJ, Zanca C, Bi J, Ikegami S, Cahill GL, et al. An LXR-Cholesterol Axis Creates a Metabolic Co-Dependency for Brain Cancers. Cancer Cell . 2016 Nov;30(5):683–93. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S1535610816304433
- [43] Giannarelli C, Cimmino G, Connolly TM, Ibanez B, Garcia Ruiz JM, Alique M, et al. Synergistic effect of liver X receptor activation and simvastatin on plaque regression and stabilization: an magnetic resonance imaging study in a model of advanced atherosclerosis. Eur Heart J . 2012 Jan;33(2):264–73. Available from: https://academic.oup.com/eurheartj/articlelookup/doi/10.1093/eurheartj/ehr136
- [44] Cannon M V., Yu H, Candido WM, Dokter MM, Lindstedt E-L, Silljé HHW, et al. The liver X receptor agonist AZ876 protects against pathological cardiac hypertrophy and fibrosis without lipogenic side effects. Eur J Heart Fail . 2015 Mar;17(3):273–82. Available from: https://onlinelibrary.wiley.com/doi/10.1002 /ejhf.243
- [45] van der Hoorn J, Lindén D, Lindahl U, Bekkers M, Voskuilen M, Nilsson R, et al. Low dose of the liver X receptor agonist, AZ876, reduces atherosclerosis in APOE\*3Leiden mice without affecting liver or plasma triglyceride levels. Br J Pharmacol . 2011 Apr;162(7):1553–63. Available from:

https://onlinelibrary.wiley.com/doi/10.1111 /j.1476-5381.2010.01168.x

- [46] Kirchgessner TG, Martin R, Sleph P, Grimm D, Liu X, Lupisella J, et al. Pharmacological Characterization of a Novel Liver X Receptor Agonist with Partial LXR a Activity and a Favorable Window in Nonhuman Primates. J Pharmacol Exp Ther 2015 Feb;352(2):305-14. Available from: http://jpet.aspetjournals.org/lookup/doi/10. 1124/jpet.114.219923
- [47] Kick E, Martin R, Xie Y, Flatt B, Schweiger E, Wang T-L, et al. Liver X Receptor (LXR) partial agonists: Biaryl pyrazoles and imidazoles displaying a preference for LXRβ. Bioorg Med Chem Lett . 2015 Jan;25(2):372–7. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0960894X1401213X
- [48] Kuipers I, Li J, Vreeswijk-Baudoin I, Koster J, van der Harst P, Silljé HHW, et al. Activation of liver X receptors with T0901317 attenuates cardiac hypertrophy in vivo. Eur J Heart Fail . 2010 Oct;12(10):1042–50. Available from: http://doi.wiley.com/10.1093/eurjhf/hfq109
- [49] Kirchgessner TG, Sleph P, Ostrowski J, Lupisella J, Ryan CS, Liu X, et al. Beneficial and Adverse Effects of an LXR Agonist on Human Lipid and Lipoprotein Metabolism and Circulating Neutrophils. Cell Metab . 2016 Aug;24(2):223–33. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S155041311630362X
- [50] Morello F. Liver X receptors and regulate renin expression in vivo. J Clin Invest . 2005

Jul 1;115(7):1913–22. Available from: http://www.jci.org/cgi/doi/10.1172/JCI245 94

- [51] Tamura K, Chen YE, Horiuchi M, Chen Q, Daviet L, Yang Z, et al. LXRα functions as a cAMP-responsive transcriptional regulator of gene expression. Proc Natl Acad Sci . 2000 Jul 18;97(15):8513–8. Available from: https://pnas.org/doi/full/10.1073/pnas.1005 19097
- [52] Cannon M V, Silljé HH, Sijbesma JW, Vreeswijk-Baudoin I, Ciapaite J, van der Sluis B, et al. Cardiac <scp>LXR</scp> α protects against pathological cardiac hypertrophy and dysfunction by enhancing glucose uptake and utilization. EMBO Mol Med . 2015 Sep 14;7(9):1229–43. Available from: https://onlinelibrary.wiley.com/doi/10.1525

2/emmm.201404669

- [53] Imayama I, Ichiki T, Patton D, Inanaga K, Miyazaki R, Ohtsubo H, et al. Liver X Receptor Activator Downregulates Angiotensin II Type 1 Receptor Expression Through Dephosphorylation of Sp1. Hypertension . 2008 Jun;51(6):1631–6. Available from: https://www.ahajournals.org/doi/10.1161/H YPERTENSIONAHA.107.106963
- [54] Leik CE, Carson NL, Hennan JK, Basso MD, Liu Q-Y, Crandall DL, et al. GW3965, a synthetic liver X receptor (LXR) agonist, reduces angiotensin II-mediated pressor responses in Sprague-Dawley rats. Br J Pharmacol . 2007 Jun;151(4):450–6. Available from:

http://doi.wiley.com/10.1038/sj.bjp.070724 1

[55] Han S, Bal NB, Sadi G, Usanmaz SE,Uludag MO, Demirel-Yilmaz E. The effectsof LXR agonist GW3965 on vascular

reactivity and inflammation in hypertensive rat aorta. Life Sci . 2018 Nov;213:287–93. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0024320518306672